

Fundamentals of enzyme kinetics and thermodynamic analysis for microbial communities

Karel Olavarria

Delft University of Technology

kogamez@gmail.com

Today:

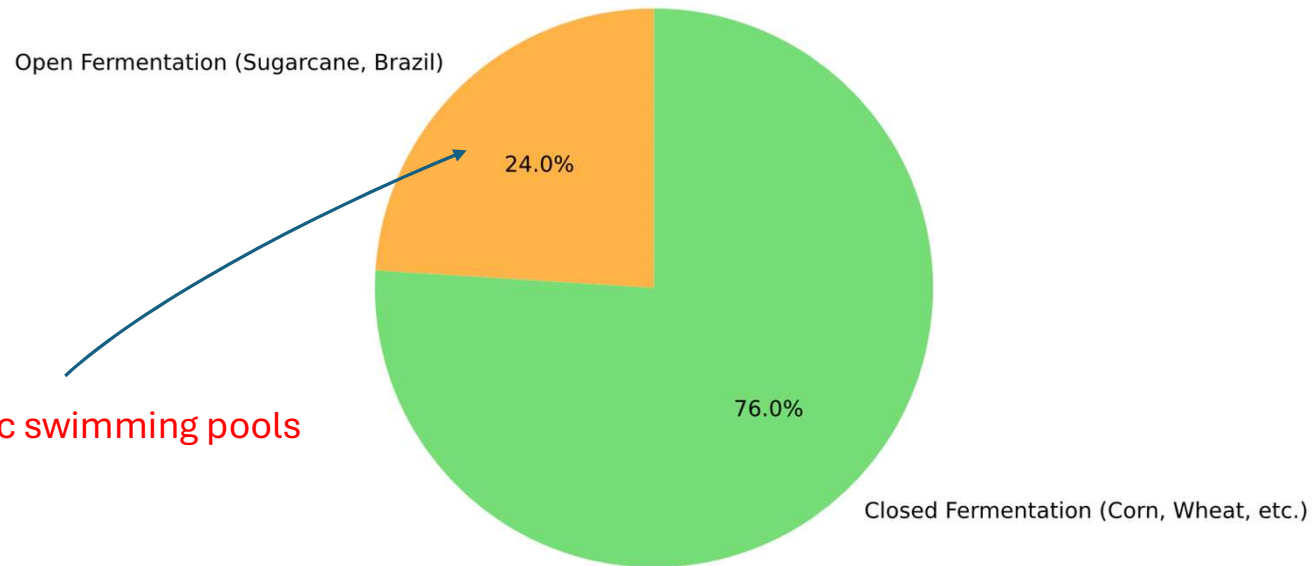
- 10:00 – 13:00 Djordje and Timmy – theoretical models in microbial ecology.
- **14:00 - 14:45 *Invited Lecture* –enzyme kinetics for microbial communities**
- **15:00 – 18:00 Flux Balance Analysis: Timmy**

Tomorrow:

- **9:00 – 11:30 Thermodynamics and computational practice**
- 12:00 – 13:00 *Invited lecture* – Djordje. Dynamic metabolic models
- 14:00 – 18:00 Djordje – Dynamic FBA y Resource allocation.

The size of the problem we are going to study today

Global Bioethanol Production: 120–125 billion liters (2024)

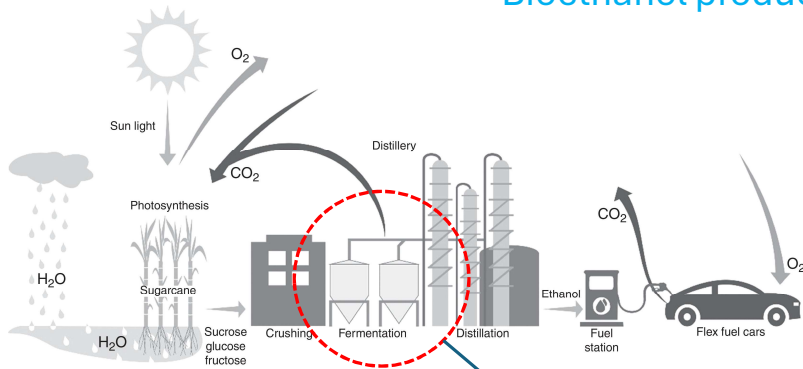


12000 Olympic swimming pools

Bioethanol in Brazil is mainly produced in open fermentation

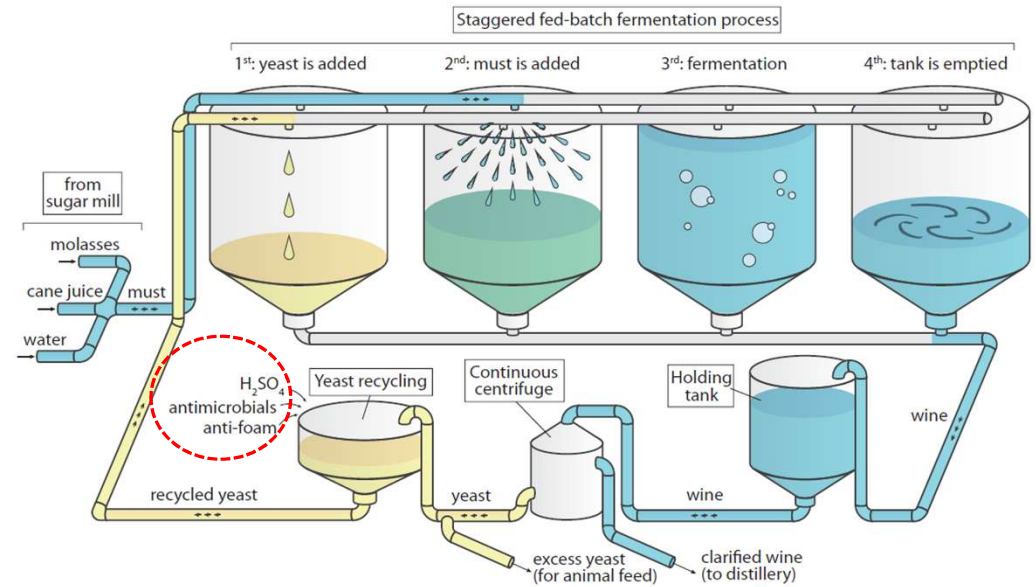


Bioethanol production, using open fermentations, in a nutshell



Source: <https://doi.org/10.1016/j.bjm.2016.10.003>

(a) Bioethanol production with yeast recycling



Source: <https://doi.org/10.1093/g3journal/jkad104>

Challenges of the open fermentations

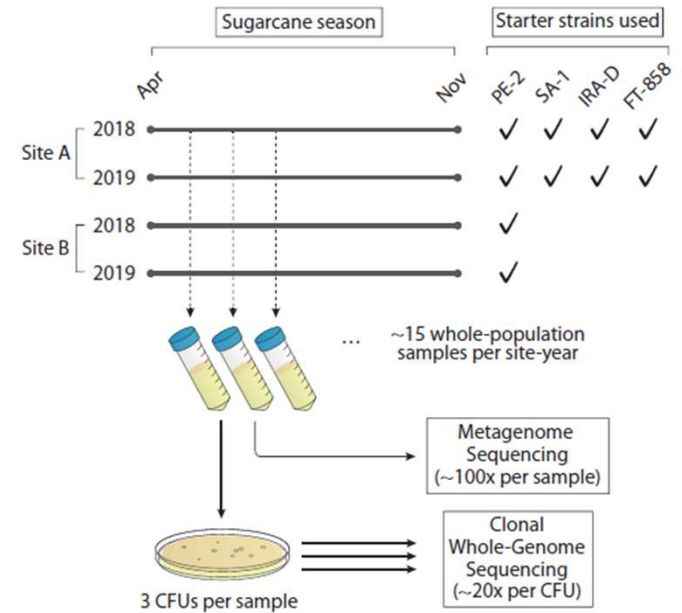


G3, 2023, 13(7), jkad104
<https://doi.org/10.1093/g3journal/jkad104>
Advance Access Publication Date: 2 June 2023
Investigation

Yeast population dynamics in Brazilian bioethanol production

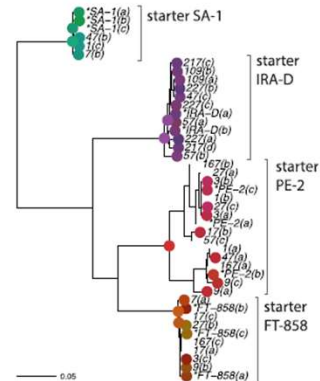
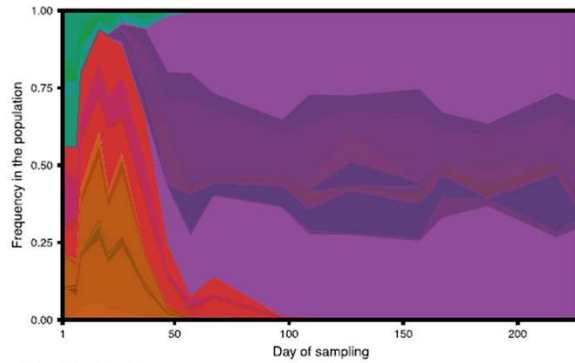
Artur Rego-Costa,^{1,†} I-Ting Huang,^{1,†} Michael M. Desai,^{1,2,3,4} Andreas K. Gombert ^{5,*}

(b) Sampling and sequencing

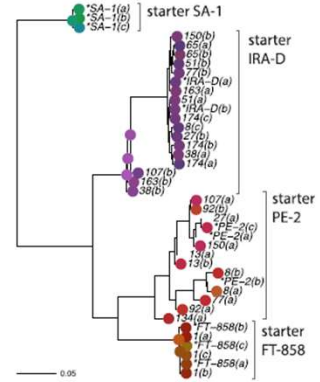
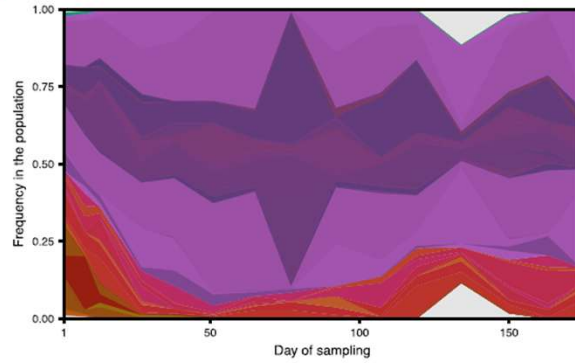


The yeast populations are dynamic

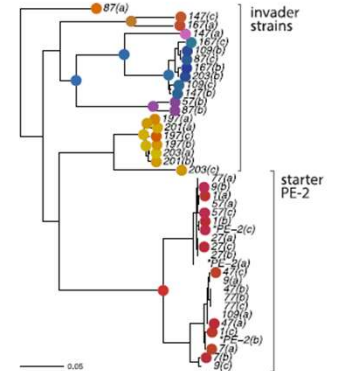
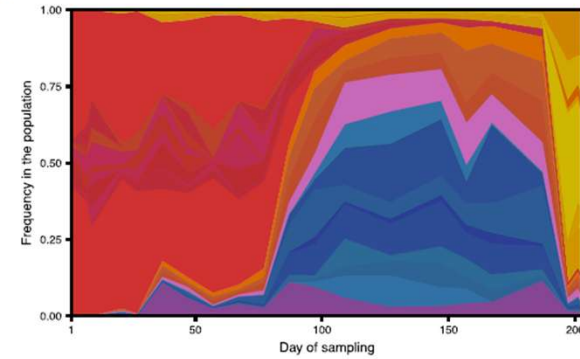
(a) Site A - 2018



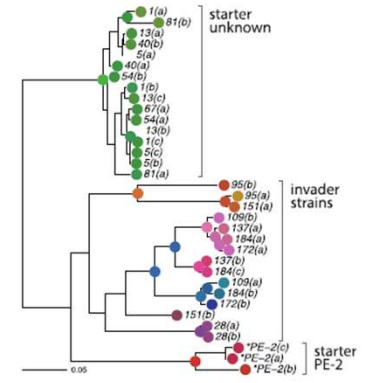
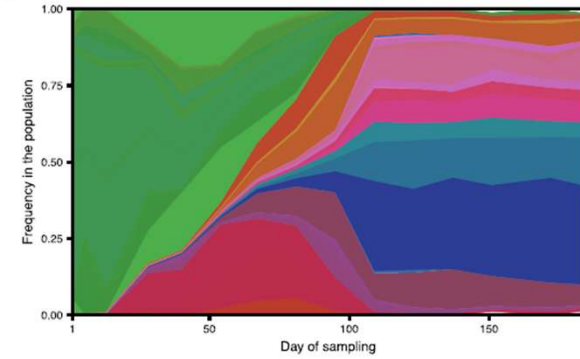
(b) Site A - 2019



(a) Site B - 2018



(b) Site B - 2019



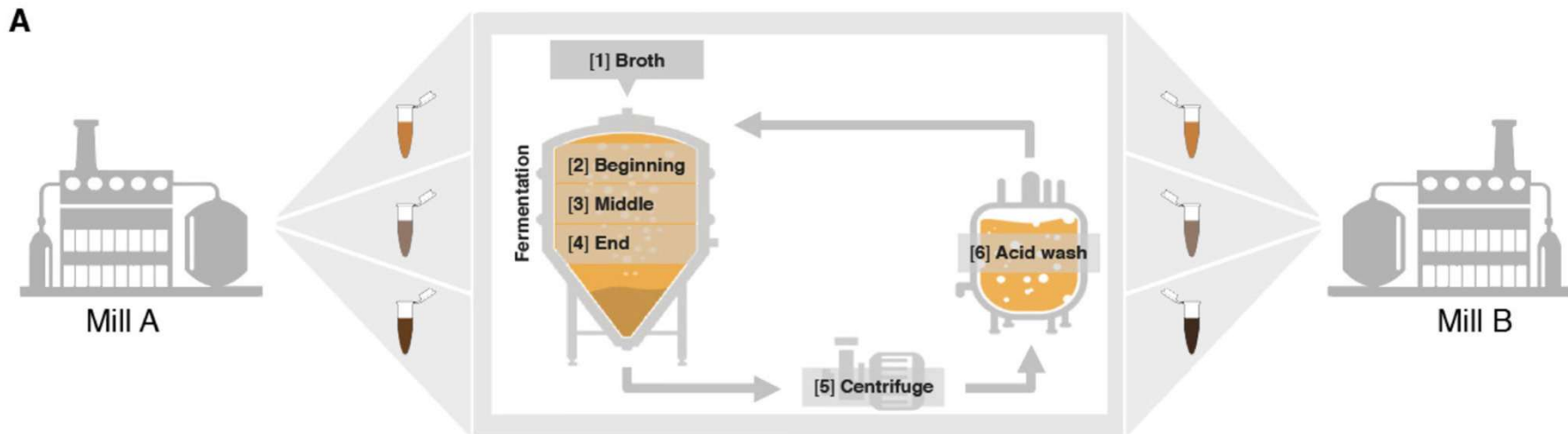
Strain dynamics of contaminating bacteria modulate the yield of ethanol biorefineries

Received: 24 August 2022

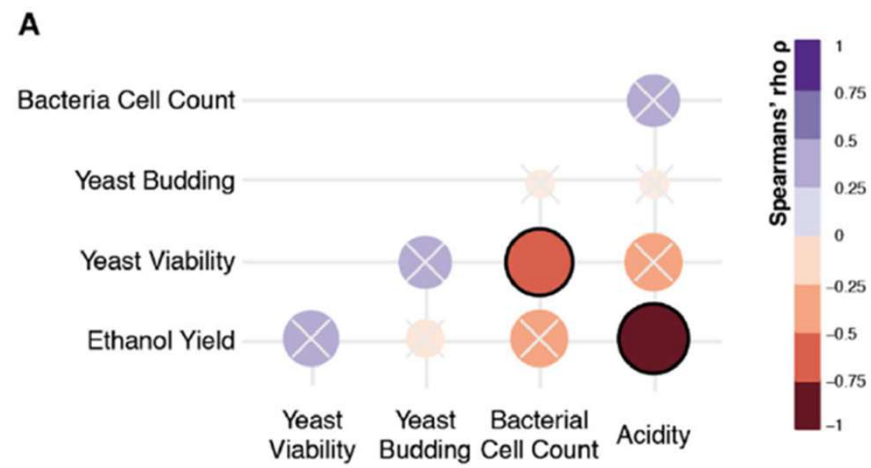
Accepted: 16 June 2024

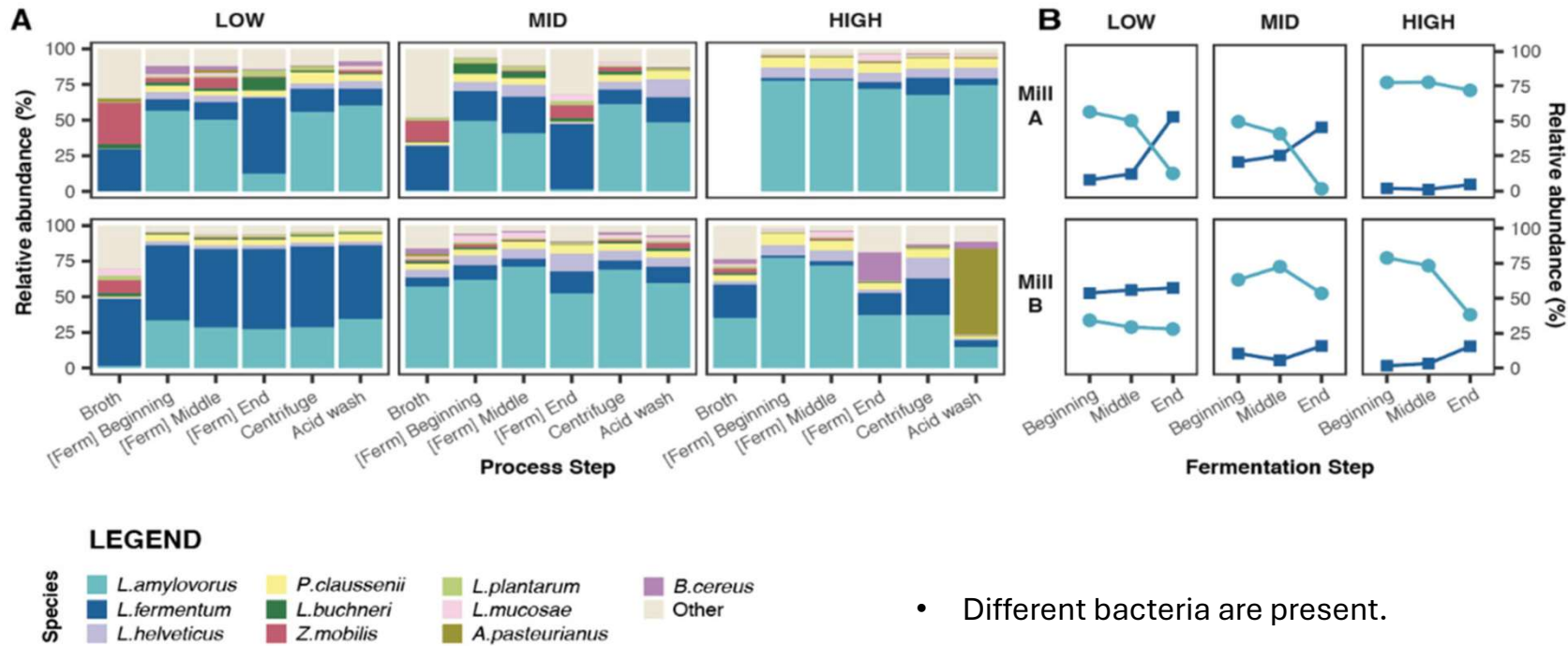
Published online: 22 June 2024

Felipe Senne de Oliveira Lino ^{1,6}, Shilpa Garg ^{1,6}, Simone S. Li^{1,2},
Maria-Anna Misiakou¹, Kang Kang ³, Bruno Labate Vale da Costa⁴,
Tobias Svend-Aage Beyer-Pedersen ¹, Thamis Guerra Giacon ⁵,
Thiago Olitta Basso ⁵, Gianni Panagiotou ³ &
Morten Otto Alexander Sommer ¹ ✉



Number of bacteria and the acids produced by them negatively affect ethanol production





- Different bacteria are present.
- *L. amylovorus* and *L. fermentum* are the dominant bacteria.
- High-performing batches contained more *L. amylovorus*.
- Across all batches and mills, *L. fermentum* increases and *L. amylovorus* decreases by the end of fermentation.

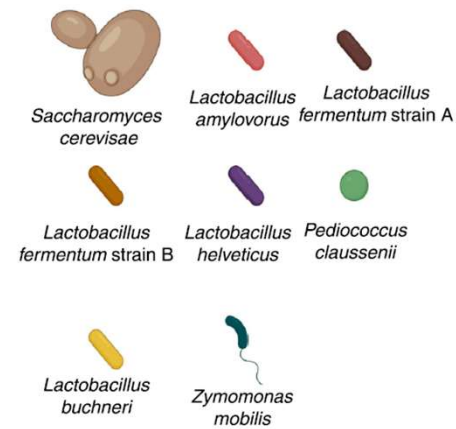
Complex yeast–bacteria interactions affect the yield of industrial ethanol fermentation

[Felipe Senne de Oliveira Lino](#), [Djordje Bajić](#), [Jean Celestin Charles Vila](#), [Alvaro Sánchez](#) & [Morten Otto](#)

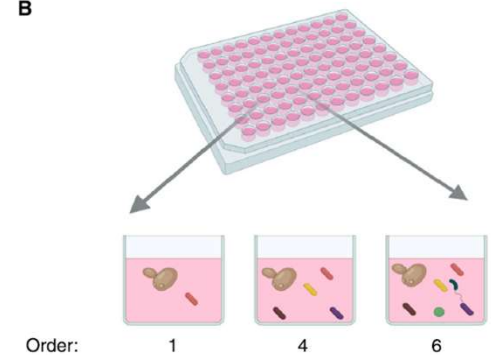
[Alexander Sommer](#) 

[Nature Communications](#) **12**, Article number: 1498 (2021) | [Cite this article](#)

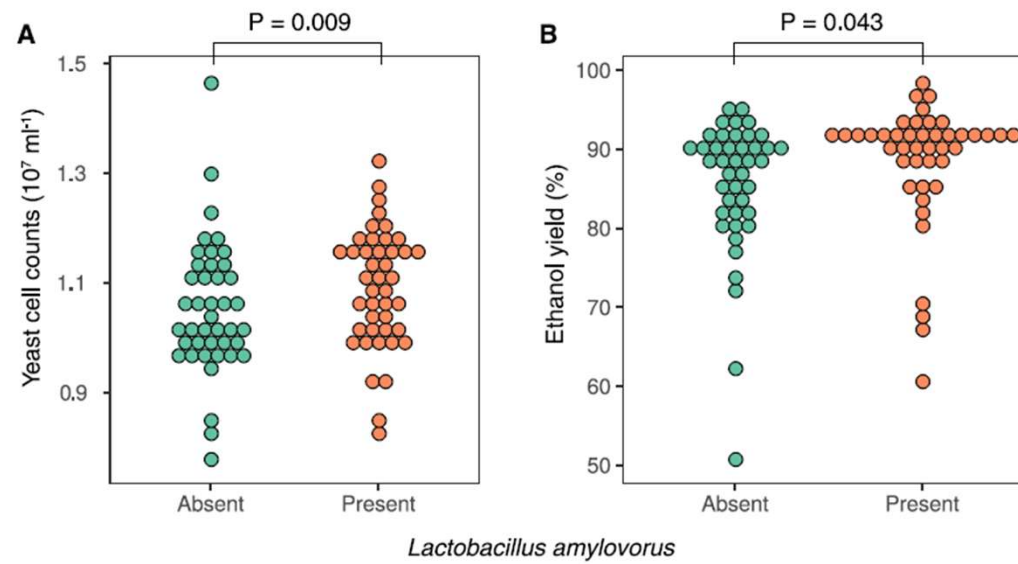
A



B



The bacterium *Lactobacillus amylovorus* has a positive effect on the ethanol production



In open fermentations:

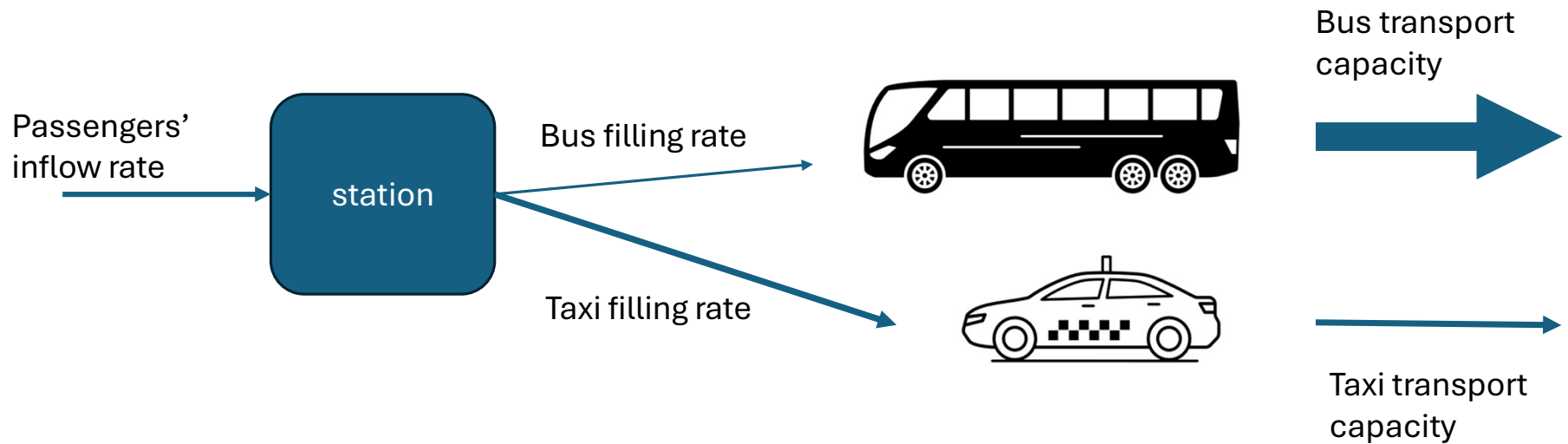
- Yeast populations are dynamic.
- Different bacteria are present and influence the ethanol production.
- Some bacteria have negative effects on the ethanol production, but some bacteria have a positive effect.

Our goals:

- Concepts from enzyme kinetics useful to study microbial communities
- Stoichiometric analysis of metabolic networks (Timmy's lecture)
- Integrating enzyme kinetics, stoichiometric analysis and thermodynamics

Let's start with a fundamental concept: **saturation**

- One bus station
- Passengers arriving to the station at different rates
- Passengers can be transported out of the station by bus or by taxi



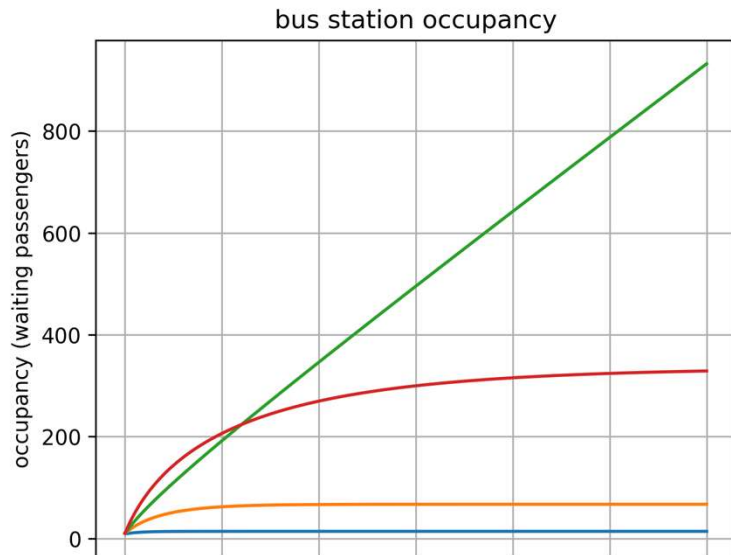
Which is the most effective way of transportation?

S : passengers waiting at the station
E : empty vehicles ready to be filled
ES : vehicles currently being filled with passengers
P : cumulative passengers already transported

k1 : rate at which passengers enter vehicles
k1r : rate at which passengers leave vehicles (before departure)
k2 : rate at which filled vehicles depart
f : external passenger influx (new arrivals per minute)

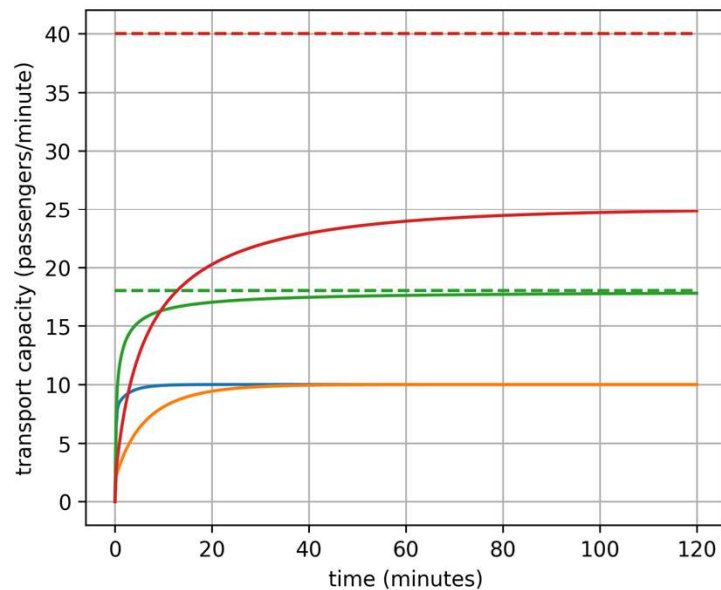
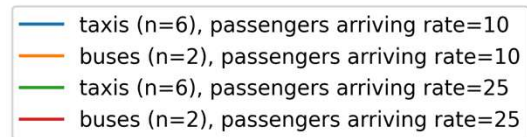
taxis are easy to fill but small capacity
buses are harder to fill but large capacity

$dS = f - k1 \cdot S \cdot E + k1r \cdot ES$ # passengers at the station
 $dE = -k1 \cdot S \cdot E + (k1r + k2) \cdot ES$ # empty vehicles
 $dES = k1 \cdot S \cdot E - (k1r + k2) \cdot ES$ # filled vehicles
 $dP = k2 \cdot ES$ # passengers transported out of the station



Set:

- Taxis carry 3 passengers
- Buses carry 20 passengers



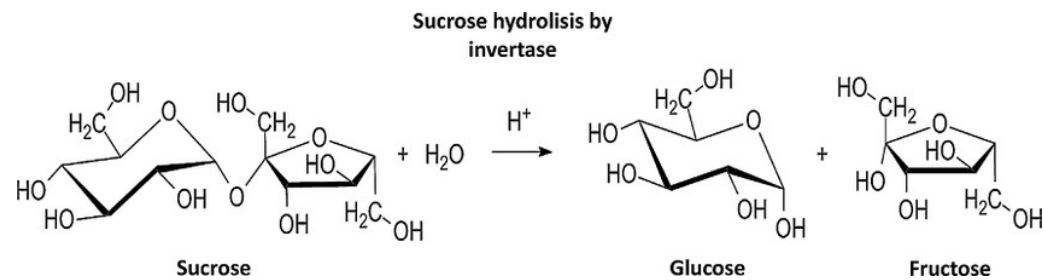
- **What are representing the dashed lines?**
- What happens with the transportation by **taxi** when the passenger arriving rate is **25 passengers/minute**?



Maud Menten

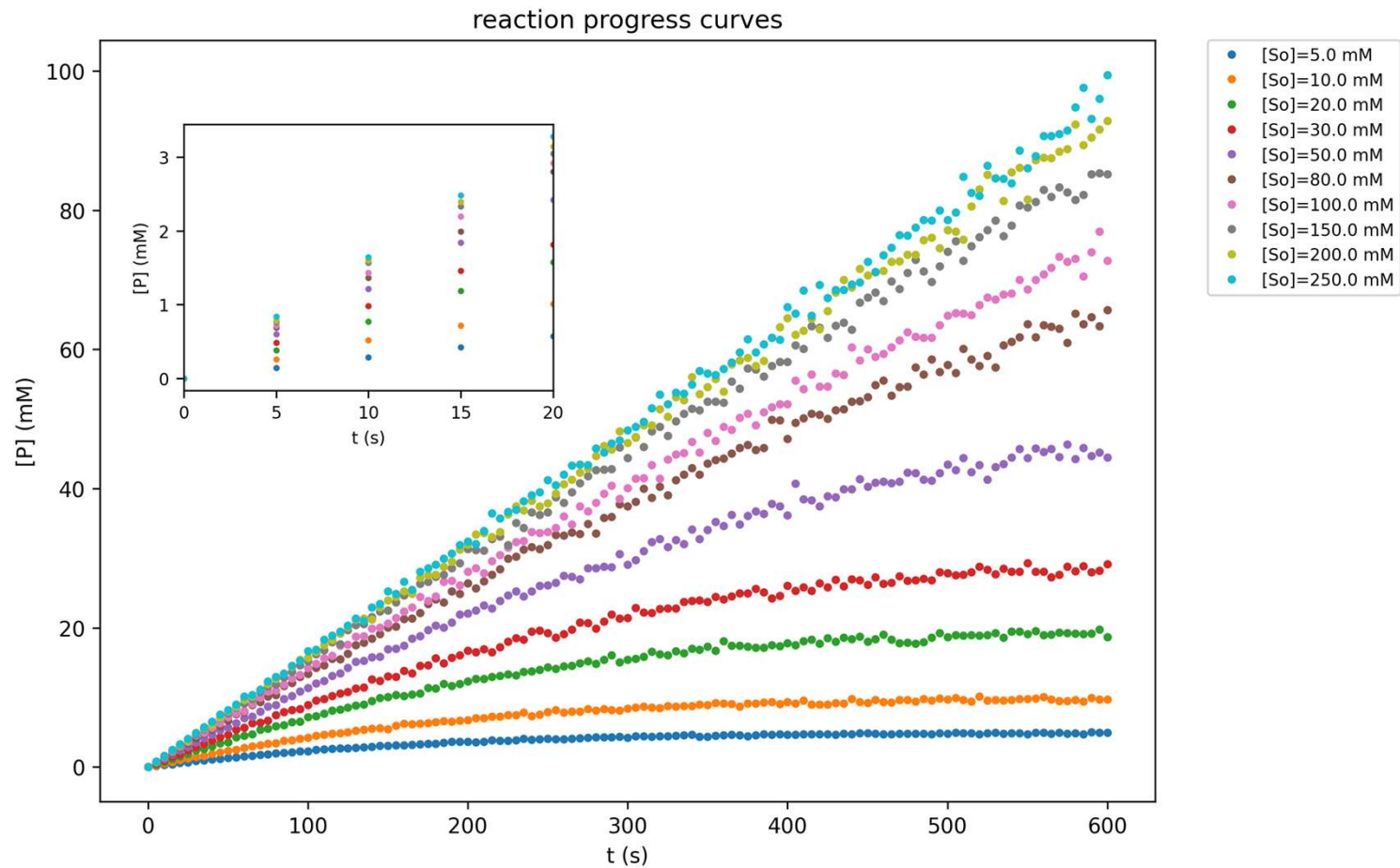
Leonor Michaelis

Michaelis, L., and Menten, M. (**1913**) Die kinetik der invertinwirkung, *Biochemistry Zeitung* 49, 333-369.



Happy coincidence! Sucrose is the most abundant sugar in the sugarcane juice

Michaelis and Menten also observed the phenomenon of saturation while studying the reaction catalyzed by the invertase





If there is one liter of a sucrose solution of 5 grams/liter, and we add 0.001 grams of invertase from yeast, how long it takes to hydrolyze half of the sucrose?

What do we need to know to answer this question?

Goal: predict the product formation rate in an enzyme-catalyzed reaction



E: concentration of free enzyme

S: concentration of sucrose

ES: concentration of the enzyme-substrate complex

They assumed that a **rapid equilibrium** between the species E, S and ES is established:

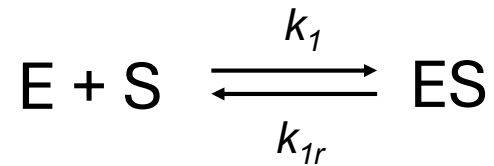
$$k_1 \gg k_2 \quad k_{1r} \gg k_2$$

They assumed that the rate-limiting step is the product formation:

$$\text{product formation rate} = k_2 * ES$$

In 1913, no experimental evidence of the existence of ES.

Rapid equilibrium model



$$rate^{ES \text{ complex formation}} = rate^{ES \text{ complex dissociation}}$$

$$k_1 * \mathbf{E(t)} * S(t) = k_{1r} * \mathbf{ES(t)}$$

Difficult to measure

$$K_S = \frac{k_{1r}}{k_1}$$

$$K_S = \frac{E(t) * S(t)}{ES(t)}$$

$$ES(t) = \frac{E(t) * S(t)}{K_S} = \frac{(E_{added} - ES(t)) * S(t)}{K_S}$$

$$ES(t) = \frac{E_{added} * S(t)}{K_S + S(t)}$$

$$product \text{ formation rate} = \frac{k_2 * E_{added} * S(t)}{K_S + S(t)} = \frac{V^{max} * S(t)}{K_M + S(t)}$$

If there is one liter of a sucrose solution of 5 grams/liter, and we add 0.001 grams of invertase from yeast, **how long** it takes to hydrolyze **half** of the sucrose?

$$\text{Product formation rate} = \frac{k_{cat} * E_{added} * S(t)}{K_M + S(t)}$$

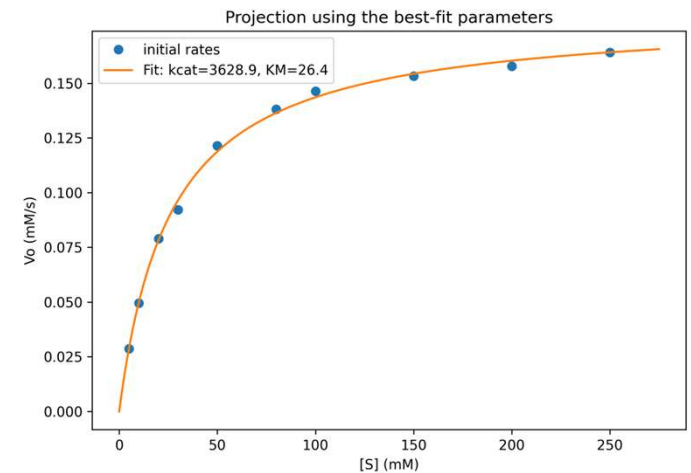
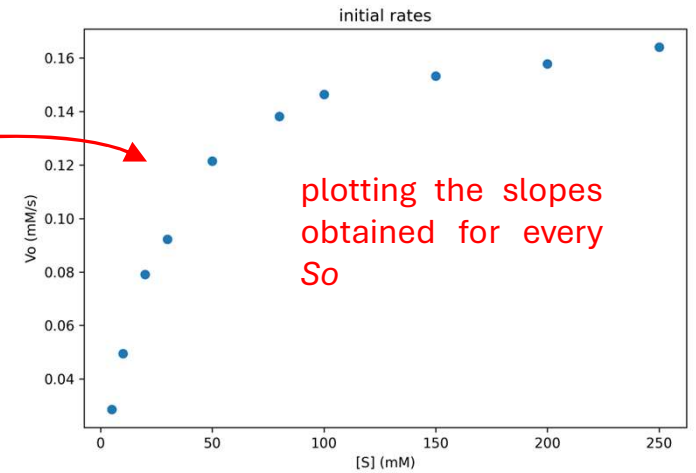
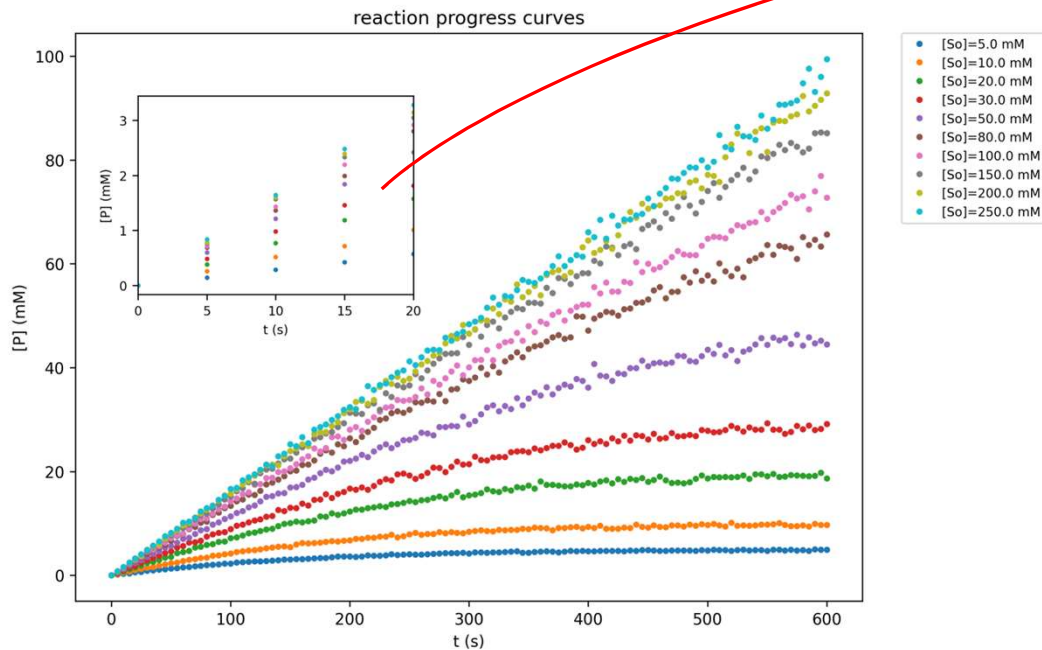
We know $S_0 = 5$ g/L, $E_{added} = 0.001$ g/L. However,

- How can we determine the kinetic parameters k_{cat} and K_M ?
- S is changing in time.
- Where is the variable **time**?

How can we determine the kinetic parameters k_{cat} and K_M ?

At the beginning of the reaction, S is *nearly* constant

→ $initial\ rate\ (V_o) = \frac{k_{cat} * E_{added} * S_o}{K_M + S_o}$



- **S is changing in time.** When half of the sucrose has been consumed, we are far from the initial rate conditions
- Where is the variable ***time***?

~~$$\text{initial rate } (V_o) = \frac{k_{cat} * E_{added} * S_o}{K_M + S_o}$$~~

$$\text{product formation rate} = \frac{dP}{dt} = -\frac{dS}{dt} = \frac{k_{cat} * E * S(t)}{K_M + S(t)}$$

$$-\frac{dS}{dt} = \frac{k_{cat} * E * S(t)}{K_M + S(t)}$$

$$S(t) + K_M * \ln \frac{S(t)}{S_o} = -k_{cat} * E * t + S_o \quad \leftarrow \text{Integrated form of the Michaelis-Menten equation}$$

$$t = \frac{S(t) + K_M * \ln \frac{S(t)}{S_o} - S_o}{-k_{cat} * E}$$

$$t = \frac{S(t) + K_M * \ln \frac{S(t)}{S_0} - S_0}{-k_{cat} * E}$$

In our particular case, $S(t) = S_0/2$

$$t_{0.5} = \frac{S_0/2 + K_M * \ln \frac{1}{2} - S_0}{-k_{cat} * E}$$

For $S_0=5 \text{ g}^*/\text{L}$, $E = 0.001 \text{ g}^*/\text{L}$, $K_M = 25 \text{ mM}$ and $k_{cat} = 3600 \text{ s}^{-1}$, $t_{0.5} \approx 6 \text{ minutes}$

*Mw_sucrose = 342.3 g/mol

*Mw_invertase = 52000 g/mol

The integrated form of the Michaelis-Menten equation can also be employed to find the values of k_{cat} and K_M using reaction progress curves analysis

$$S(t) + K_M * \ln \frac{S(t)}{S_o} = -k_{cat} * E * t + S_o$$

Between 1913 and 1997:

- Numerical integration
- (Complicated) algebraic methods

$$S(t) = K_M * \omega \left(\frac{S_o}{K_M} * e^{\frac{-k_{cat} * E * t + S_o}{K_M}} \right)$$



Santiago Schnell

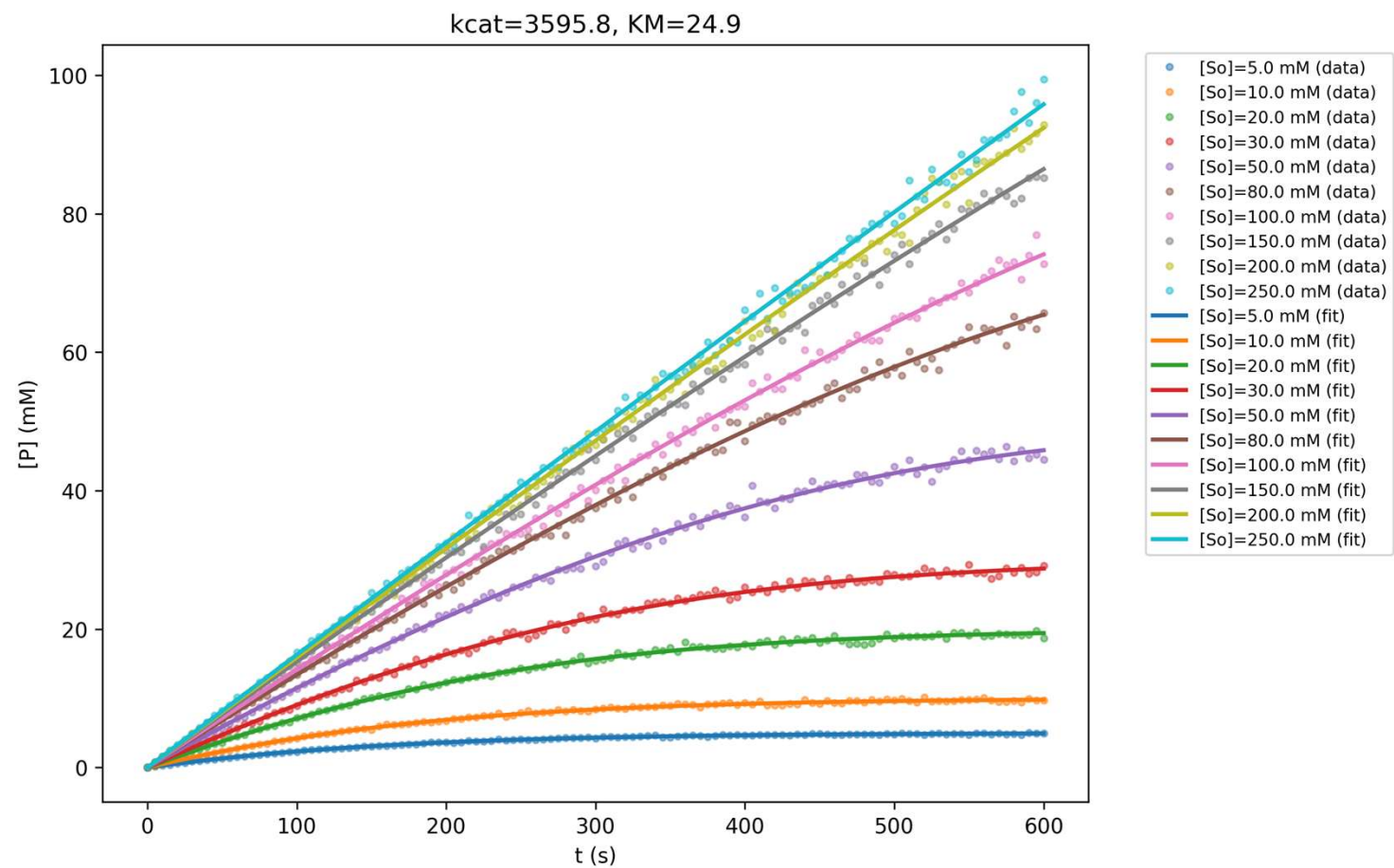


Claudio Mendoza

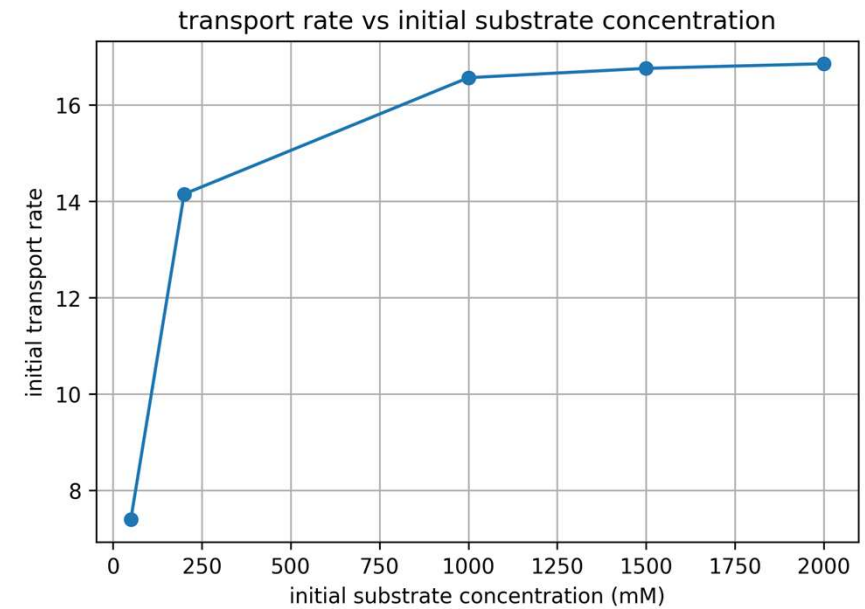
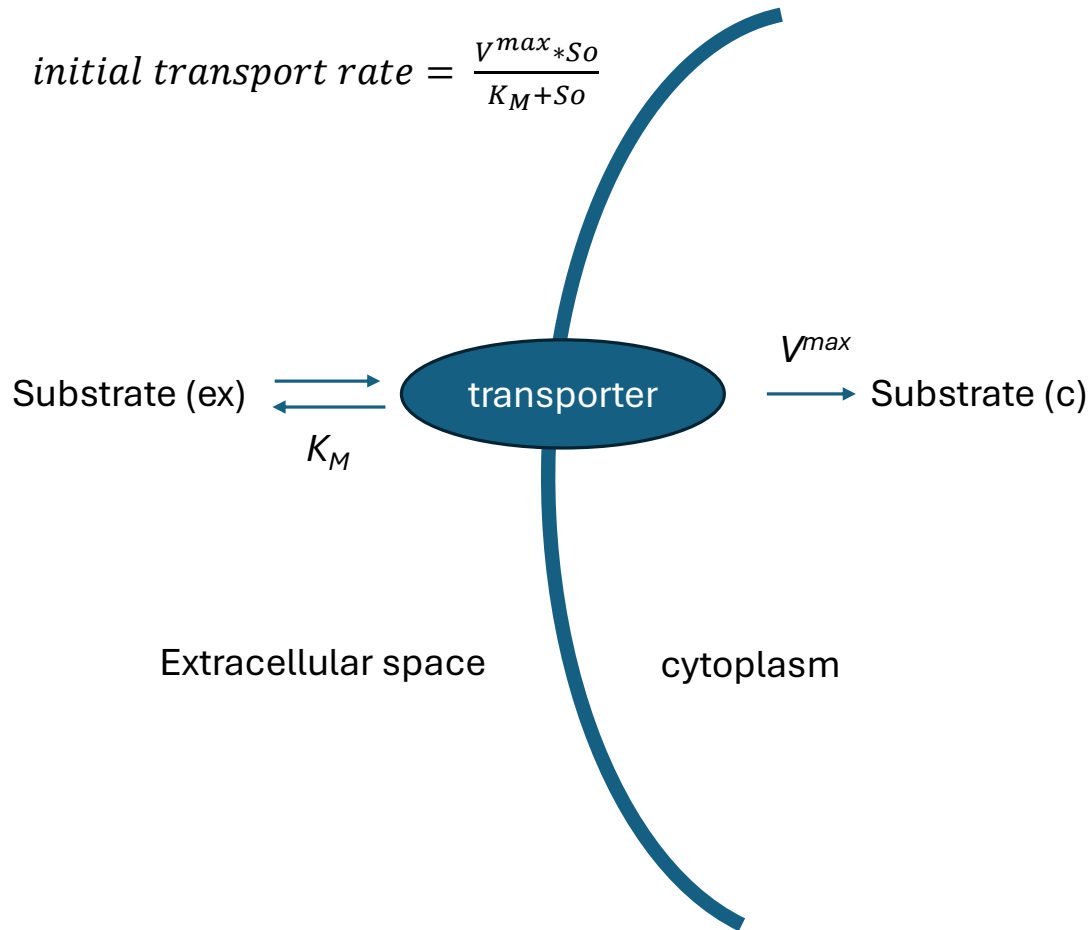


Latin got talent!

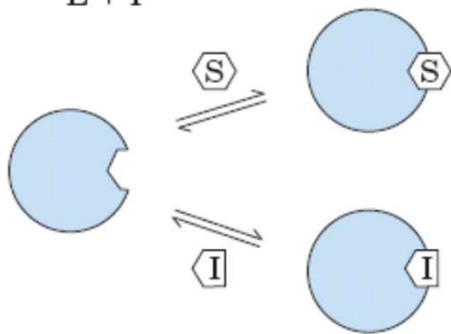
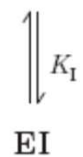
<https://doi.org/10.1006/jtbi.1997.0425>



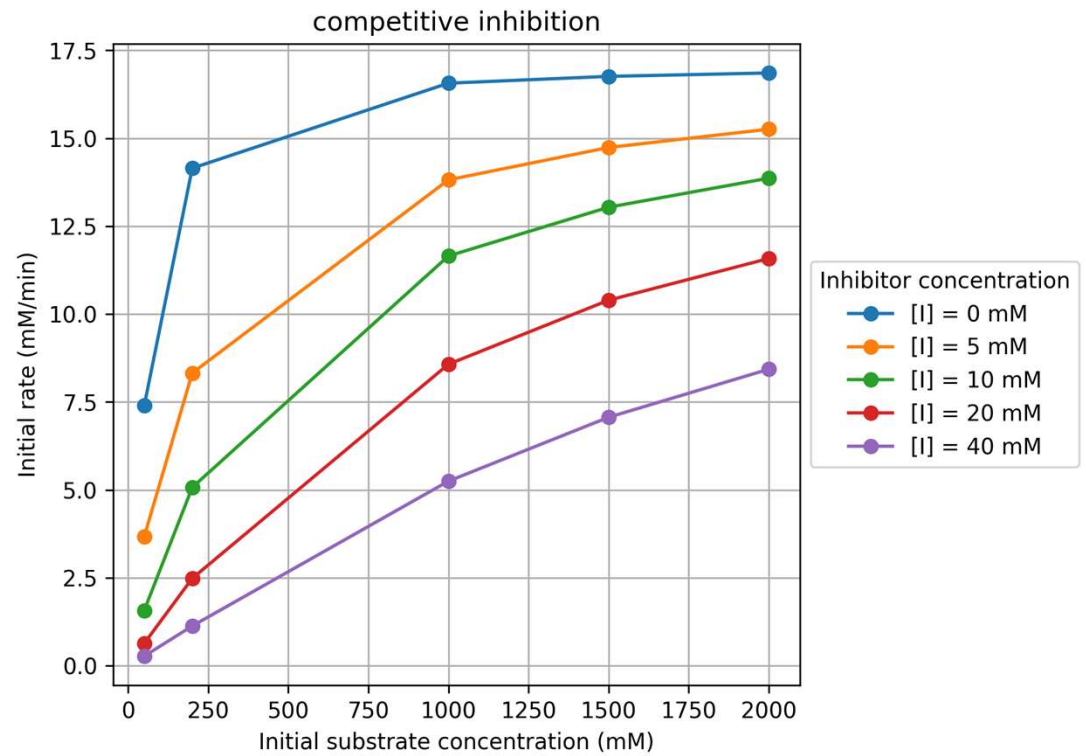
transporters can also be represented with the Michaelis-Menten equation



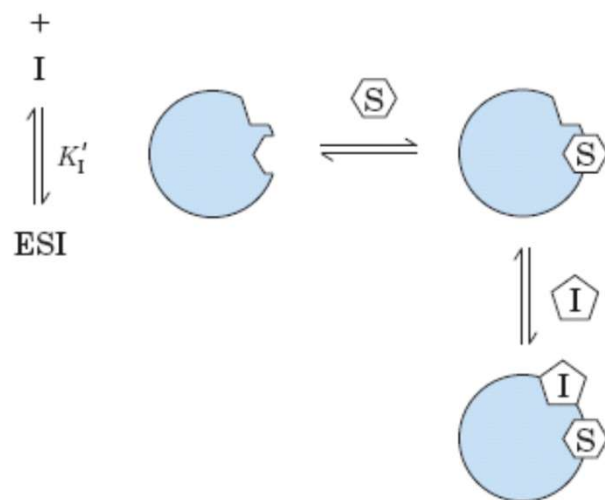
(a) Competitive inhibition



$$V_0 = \frac{V_{\max} [S]}{\alpha K_m + [S]} \quad \alpha = 1 + \frac{[I]}{K_I} \quad K_I = \frac{[E][I]}{[EI]}$$



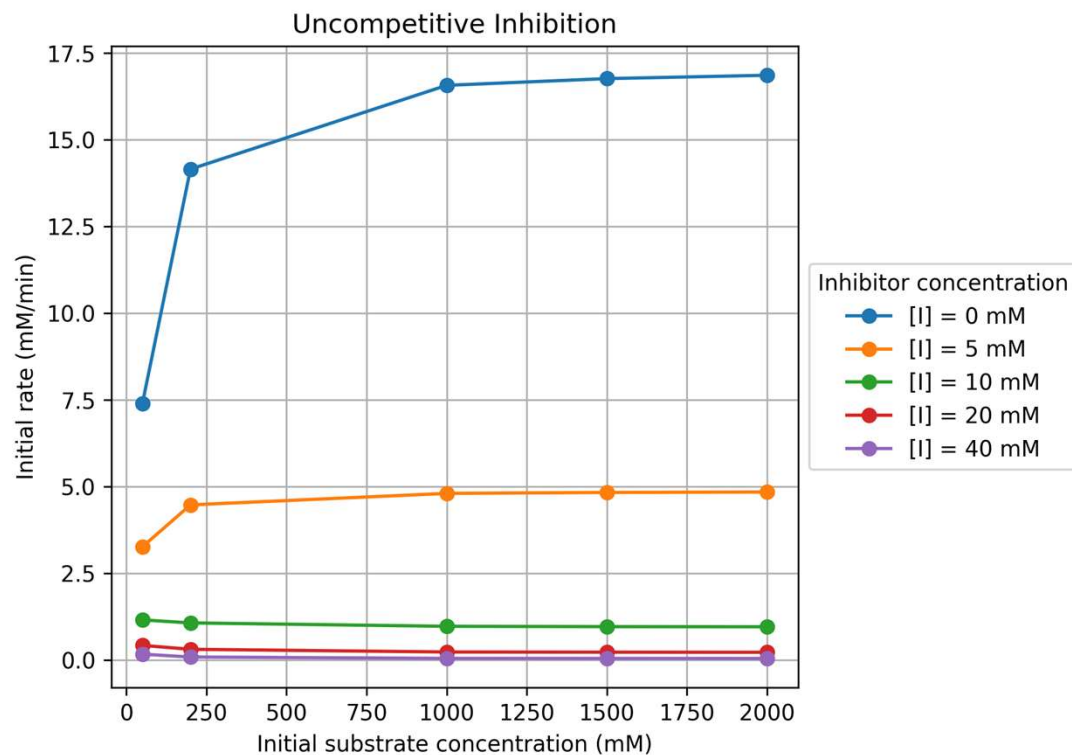
(b) Uncompetitive inhibition



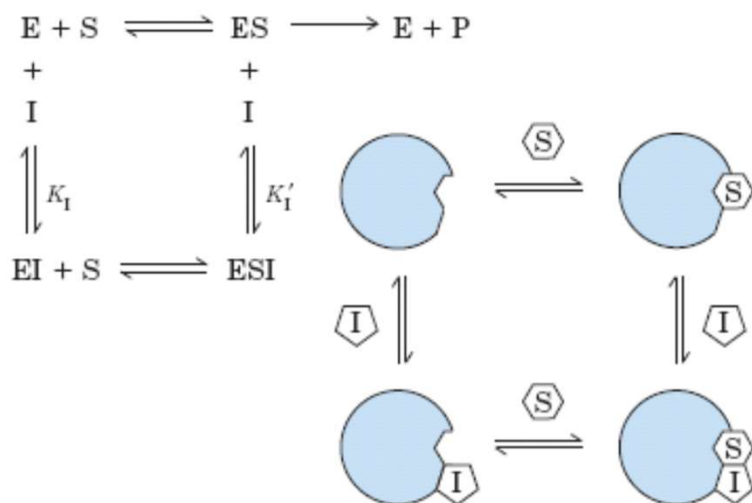
$$V_0 = \frac{V_{\max} [S]}{K_m + \alpha' [S]}$$

$$\alpha' = 1 + \frac{[I]}{K'_I}$$

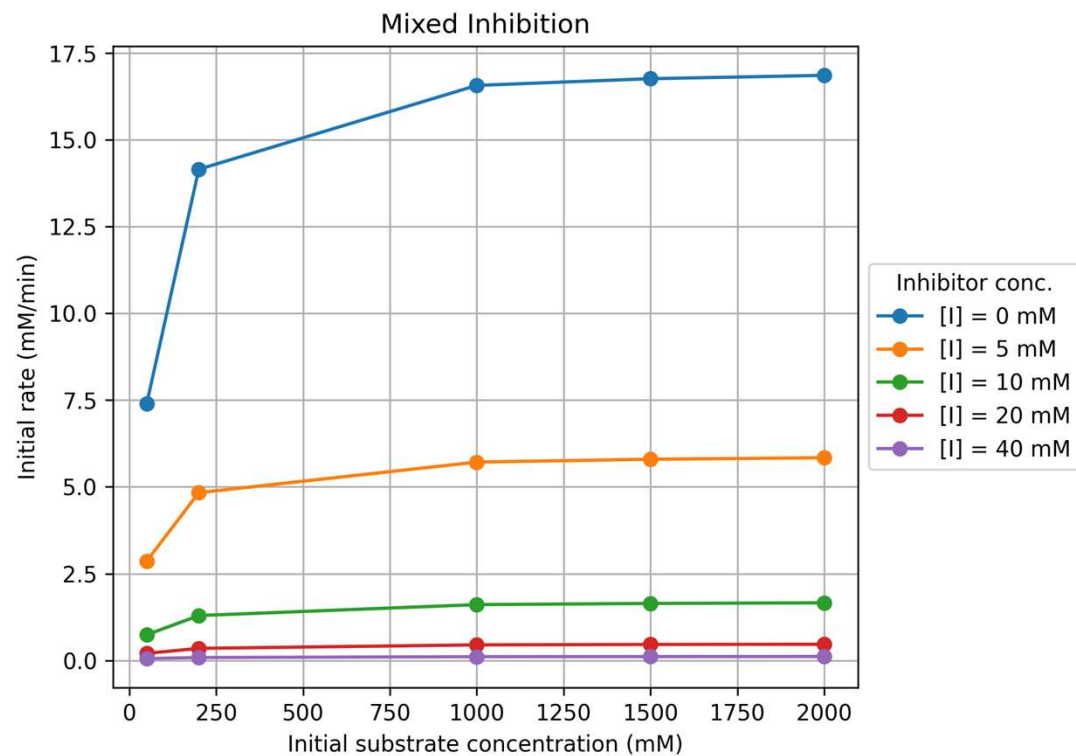
$$K'_I = \frac{[ES][I]}{[ESI]}$$



(c) Mixed inhibition



$$V_0 = \frac{V_{\max} [S]}{\alpha K_m + \alpha' [S]}$$



Coming back to our original problem:



- Sucrose is hydrolyzed by invertase (produced by yeast) into glucose and fructose via Michaelis–Menten kinetics.
- Glucose and fructose are taken up by both yeast and bacteria.
- **Assuming: Yeast transporters for glucose and fructose are inhibited by a bacterial toxin. The inhibition mechanism can be competitive or non-competitive.**

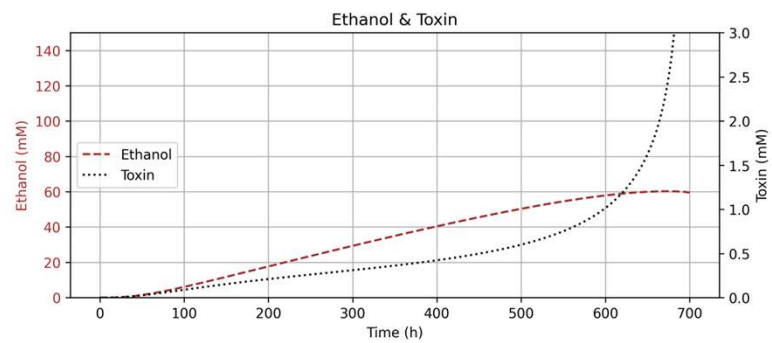
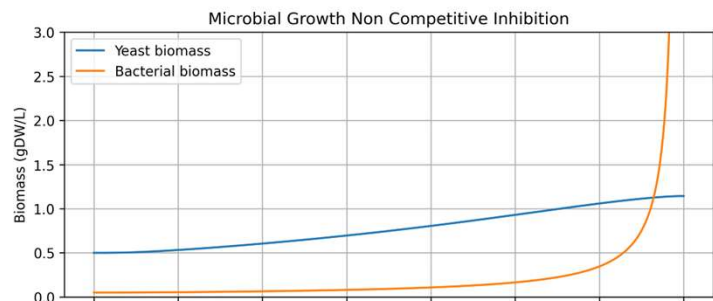
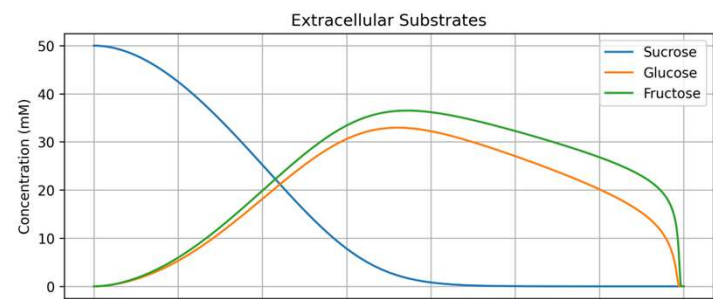
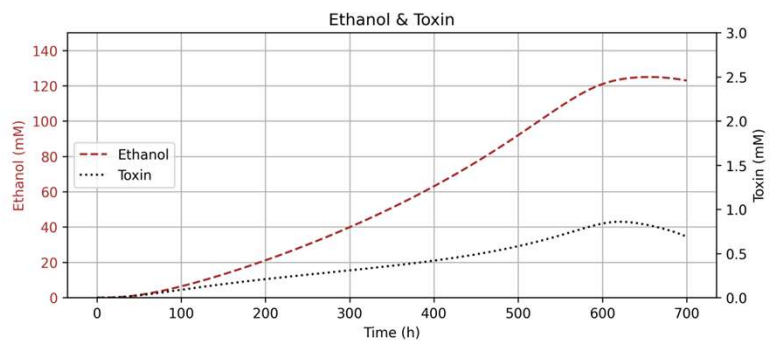
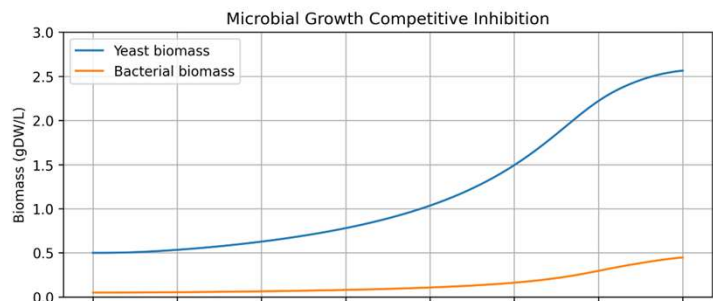
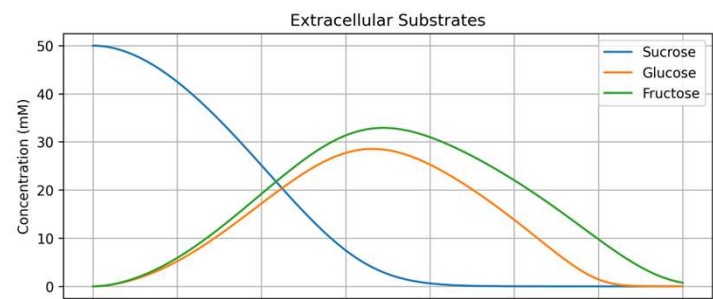


Saccharomyces cerevisiae



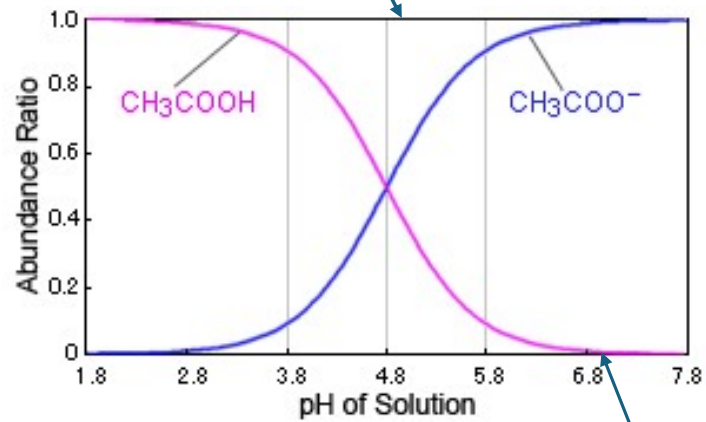
Lactobacillus fermentum strain B

Which inhibitory mechanism is more effective for the bacteria?

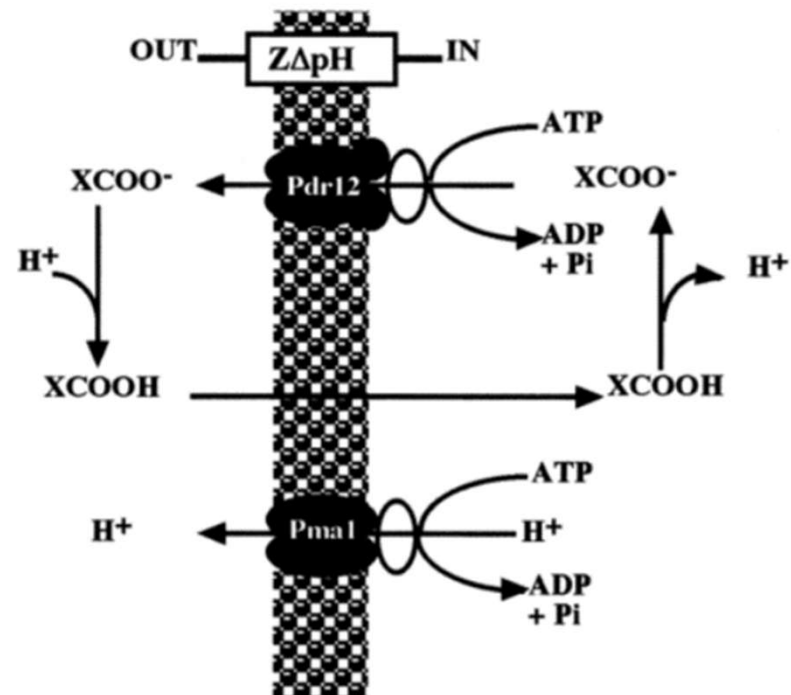


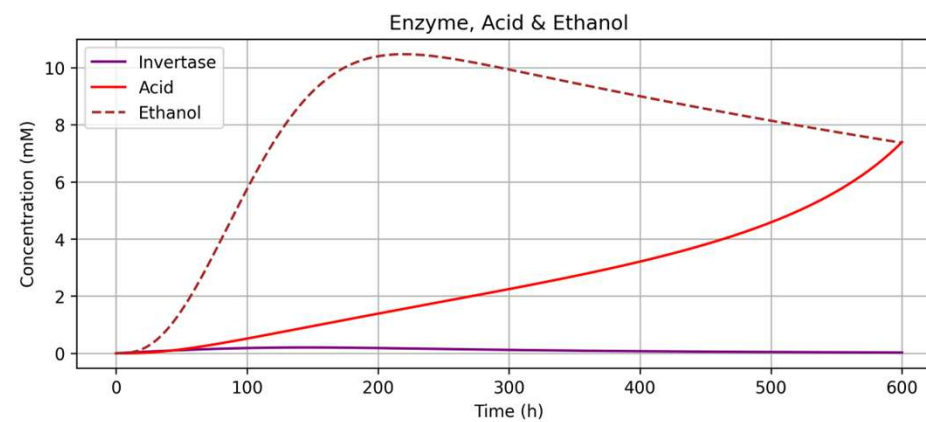
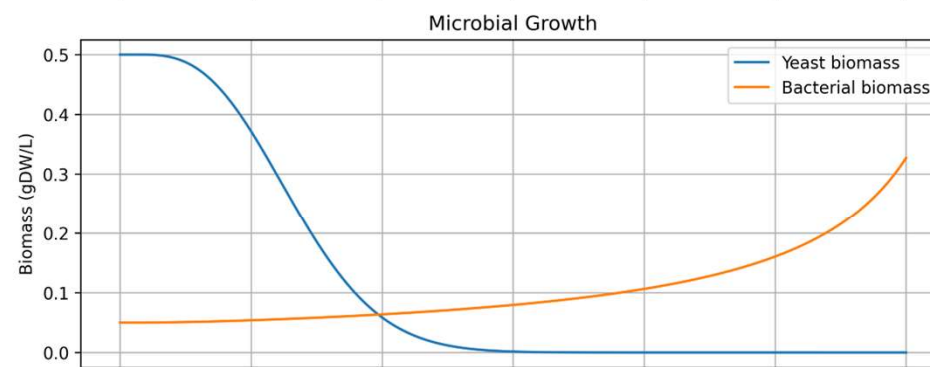
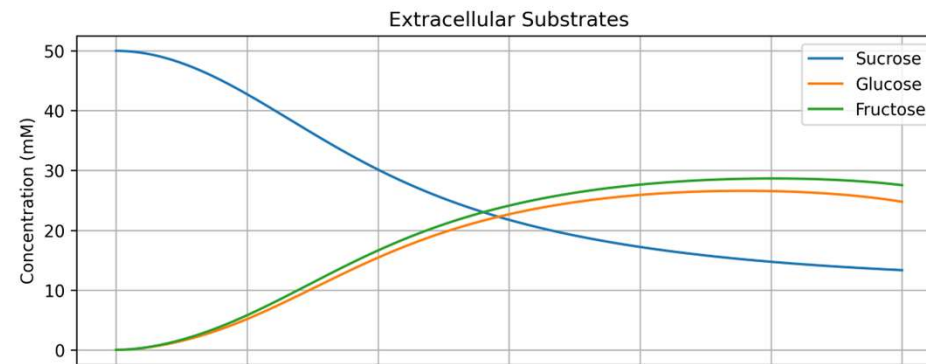
Energetic cost of the acids produced by bacteria

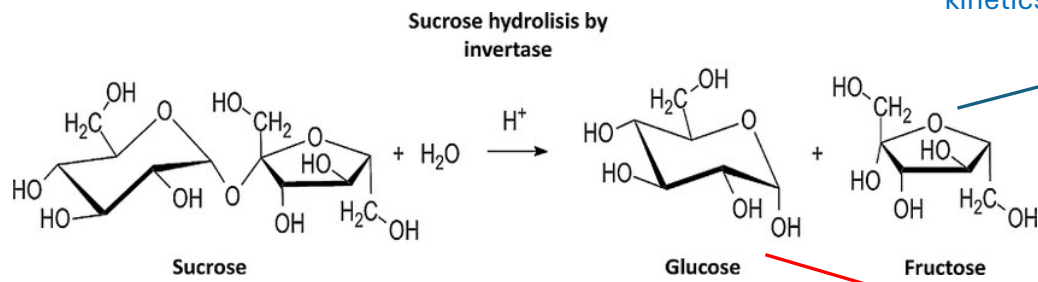
Optimal medium pH for yeast



Cytoplasmic pH







Michaelis-Menten kinetics

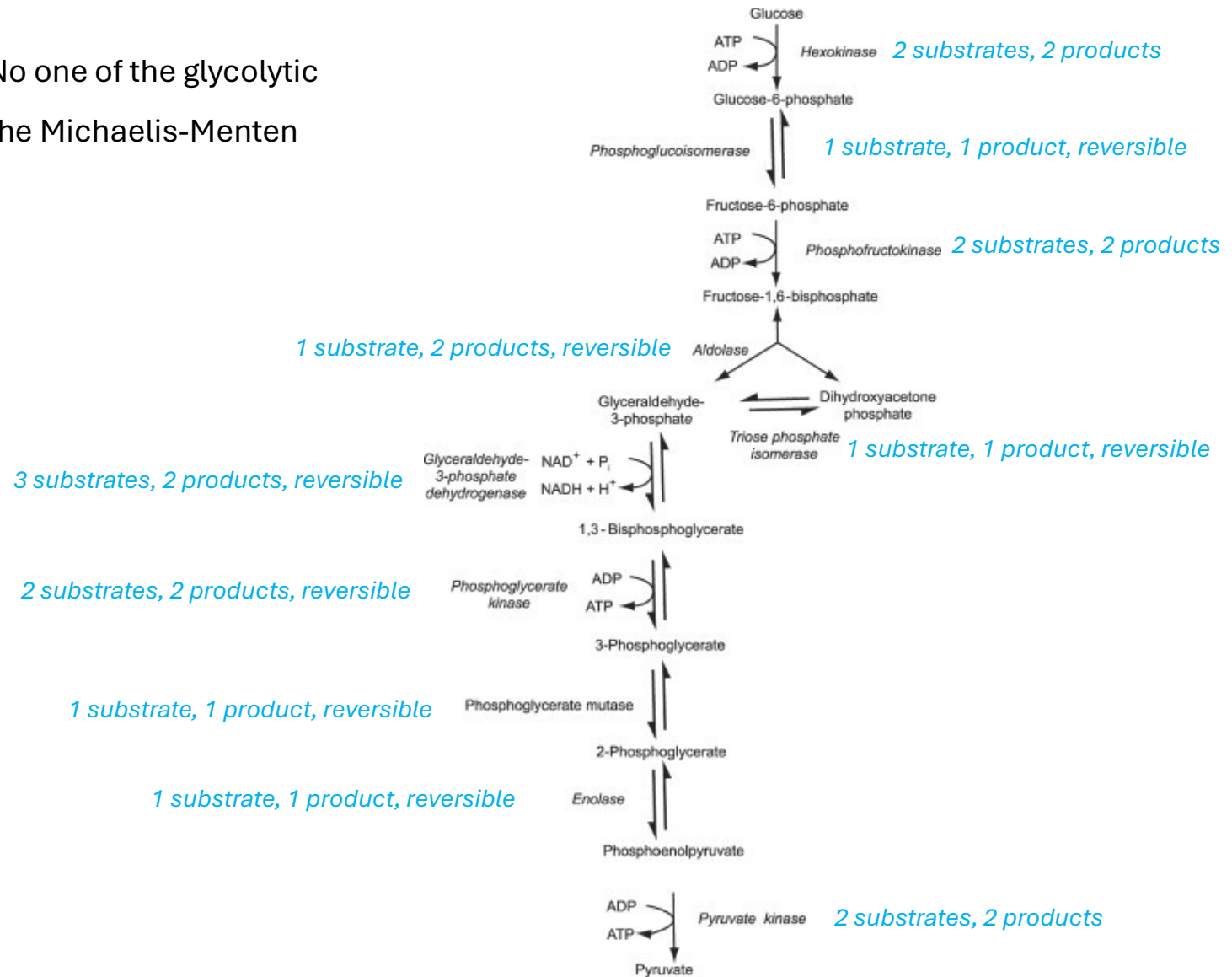
Michaelis-Menten kinetics

Michaelis-Menten kinetics

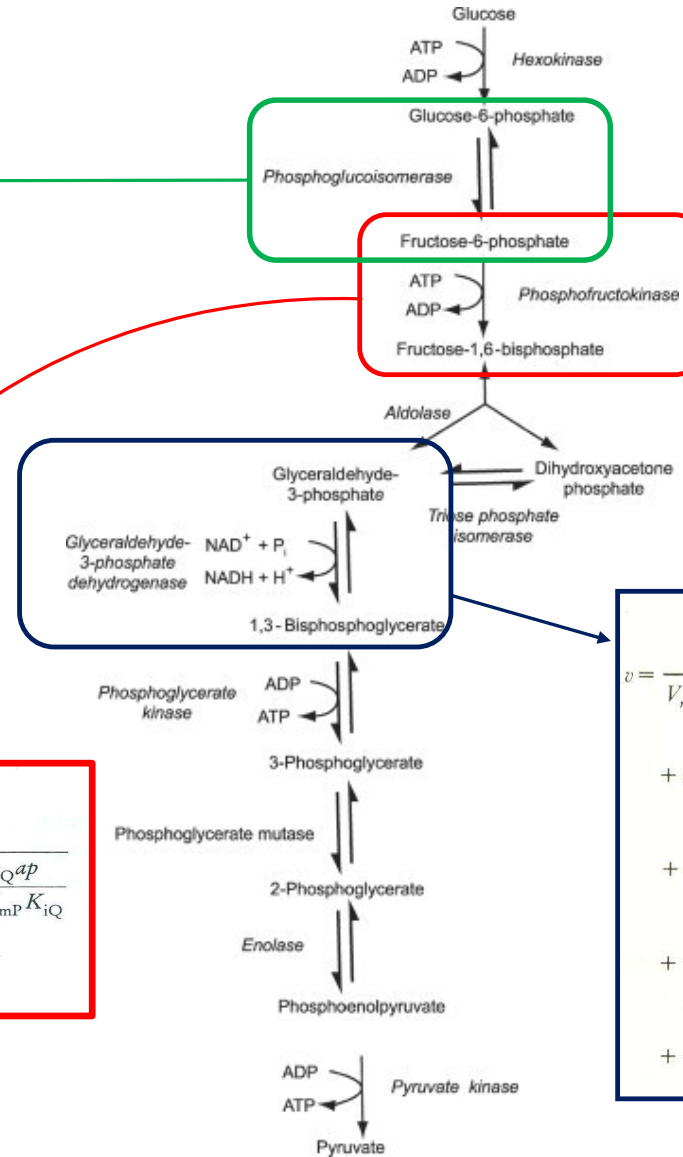
cytoplasm



Houston, we have a problem: No one of the glycolytic reaction can be described with the Michaelis-Menten equation!



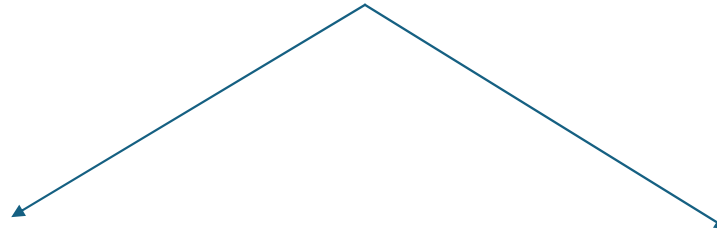
$$v = \frac{\frac{k_{cat}^f}{K_s} * E * s - \frac{k_{cat}^r}{K_p} * E * p}{1 + \frac{s}{K_s} + \frac{p}{K_p}}$$



$$v = \frac{\frac{V_{+ab}}{K_{iA}K_{mB}} - \frac{V_{-pq}}{K_{mP}K_{iQ}}}{1 + \frac{a}{K_{iA}} + \frac{K_{mA}b}{K_{iA}K_{mB}} + \frac{K_{mQ}p}{K_{mP}K_{iQ}} + \frac{q}{K_{iQ}} + \frac{ab}{K_{iA}K_{mB}} + \frac{K_{mQ}ap}{K_{iA}K_{mP}K_{iQ}} + \frac{K_{mA}bq}{K_{iA}K_{mB}K_{iQ}} + \frac{pq}{K_{mP}K_{iQ}} + \frac{abp}{K_{iA}K_{mB}K_{iP}} + \frac{bpq}{K_{iB}K_{mP}K_{iQ}}}$$

$$v = \frac{V_f V_r \left([A][B][C] - \frac{[P][Q]}{K_{eq}} \right)}{V_r K_{ia} K_{ib} K_{mC} + V_r K_{ib} K_{mC} [A] + V_r K_{ia} K_{mB} [C] + V_r K_{mC} [A][B] + K_{mB} [A][C] + V_r K_{mA} [B][C] + V_r [A][B][C] + \frac{V_f K_{mQ} [P]}{K_{eq}} + \frac{V_f K_{mP} [Q]}{K_{eq}} + \frac{V_f [P][Q]}{K_{eq}} + \frac{V_f K_{mQ} [A][P]}{K_{ia} K_{eq}} + \frac{V_f K_{mQ} [A][B][P]}{K_{ia} K_{ib} K_{eq}} + \frac{V_f K_{mQ} [A][B][C][P]}{K_{ia} K_{ib} K_{ic} K_{eq}} + \frac{V_r K_{ia} K_{mB} [C][Q]}{K_{iq}} + \frac{V_r K_{mA} [B][C][Q]}{K_{iq}} + \frac{V_r K_{ia} K_{mB} [C][P][Q]}{K_{ip} K_{iq}} + \frac{V_r K_{mA} K_{ic} [B][P][Q]}{K_{ip} K_{iq}} + \frac{V_r K_{mA} [B][C][P][Q]}{K_{ip} K_{iq}}}$$

Fortunately, there are methods to deal with this problem



Analysis of metabolic network under metabolic
steady-state (Timmy's lecture)

Integrating enzyme kinetics and thermodynamics
(second lecture)

- We reviewed some fundamental concepts of enzyme kinetics
- We learnt how some enzyme kinetics equations can help us to represent simple microbial interactions
- We saw that the actual representation of the metabolic processes can be very complex

To be continued....